

Review Article

Bullous Autoimmune Dermatoses

Clinical Features, Diagnostic Evaluation, and Treatment Options

Nina van Beek, Detlef Zillikens, Enno Schmidt

Summary

Background: Bullous autoimmune dermatoses are a clinically and immunopathologically heterogeneous group of diseases, characterized clinically by blisters or erosions of the skin and/or mucous membranes. In Germany, their prevalence is approximately 40 000 cases nationwide, and their incidence approximately 20 new cases per million people per year.

Methods: This review is based on publications that were retrieved by a selective search of the literature focusing on the current German and European guidelines.

Results: Recent years have seen the publication of guidelines, controlled prospective clinical trials, and multicenter diagnostic studies improving both diagnosis and therapy. Specific monovalent and multivariate serological test systems and pattern analysis of tissue-bound autoantibodies allow identification of the target antigens in 80–90% of patients. This enables the precise classification of disease entities, with implications for treatment selection and disease outcome. In 2019, the anti-CD20 antibody rituximab was approved by the European Medicines Agency for the treatment of moderate and severe pemphigus vulgaris, with an ensuing marked improvement in the care of the affected patients. To treat mild and moderate bullous pemphigoid, topical clobetasol propionate is recommended, in severe disease, combined with systemic treatment, i.e. usually (a) prednisolone p.o. at an initial dose of 0.5mg/kg/d, (b) an immunomodulant, e.g. dapsons or doxycycline, or (c) prednisolone plus an immunomodulant.

Conclusion: The early recognition and precise diagnostic evaluation of bullous autoimmune dermatoses now enables improved, often interdisciplinary treatment, in accordance with the available guidelines. Current research projects are focused on new treatment approaches, an improved understanding of the underlying pathophysiology, and further refinements of diagnostic techniques.

Cite this as

van Beek N, Zillikens D, Schmidt E: Bullous autoimmune dermatoses: clinical features, diagnostic evaluation, and treatment options.

Dtsch Arztebl Int 2021; 118: 413–20. DOI: 10.3238/arztebl.m2021.0136

Department of Dermatology, Venereology, and Allergology, University of Lübeck, Lübeck, Germany: Dr. med. Nina van Beek, Prof. Dr. med. Detlef Zillikens, Prof. Dr. med. Dr. rer. nat. Enno Schmidt

Lübeck Institute of Experimental Dermatology (LIED), University of Lübeck, Lübeck, Germany: Prof. Dr. med. Dr. rer. nat. Enno Schmidt

cme plus +

This article has been certified by the North Rhine Academy for Continuing Medical Education. Participation in the CME certification program is possible only over the internet: cme.aerzteblatt.de. The deadline for submissions is 17 June 2022.

Autoimmune bullous diseases (AIBD) are prototypical autoantibody-mediated autoimmune diseases in which the effects of the autoantibodies are directly visible on the skin and/or on mucous membranes. If left untreated, these diseases are potentially life-threatening due to superinfection, fluid loss, and severely restricted food intake (1–4, e1, e2).

Clinically, depending on the disease entity, vesicles, blisters, pustules, erosions, excoriations, and erythema on the skin and mucous membranes can be seen. In AIBD, autoantibodies are directed against structural proteins of the skin; in pemphigus diseases, they are directed against desmosomal proteins, which connect neighboring keratinocytes/epithelial cells, and in pemphigoid diseases, against proteins of the basement membrane zone, which connect the epidermis/epithelium and the dermis/lamina propria (*Figure 1*).

Epidemiology

The frequency of AIBD differs significantly depending on the geographic region and population evaluated (2, e3, e4). In Germany and central Europe, bullous pemphigoid is by far the most common AIBD (5, e5–e10) (*Table 1*), with an increasing incidence in recent decades (e8, e11–e13). Possible causes for the increasing incidence of bullous pemphigoid may include an aging population, the association with increasingly frequent neurological diseases and certain medications (see below), and a greater awareness of atypical variants without blistering (overview in [e4]).

The most common AIBDs in children are linear IgA dermatosis and pemphigus vulgaris (6, e14). An association with the human leukocyte antigens HLA-DRB1*04 and HLA-A*10 and a polymorphic variant in the *ST18* gene have been described for pemphigus vulgaris, while an overrepresentation of HLA-DQB1*03:01 and polymorphism in the mitochondrial *ATP8* gene has been described for bullous pemphigoid (1, 2, e3, e15, e16).

Clinical features

Pemphigus diseases

Pemphigus diseases can be classified in 4 main forms based on clinical and immunopathological features: pemphigus vulgaris, in about 70–80% of patients; pemphigus foliaceus, in about 20%; paraneoplastic pemphigus, in about 5%; and IgA pemphigus, in 1–3% (*Table 2*) (2).

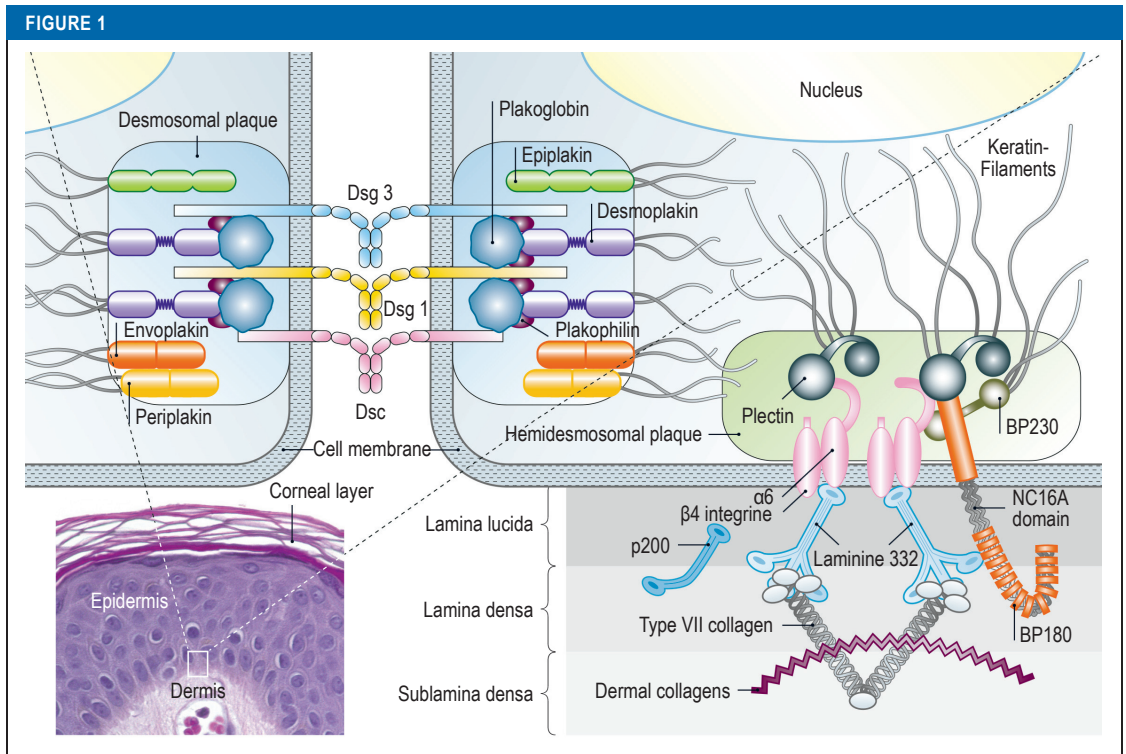


Figure 1: Schematic diagram of the autoantigens in pemphigus and pemphigoid diseases. BP180, type XVII collagen; BP230, dystonin; Dsg, desmoglein; Dsc, desmocollin

In pemphigus vulgaris, the mucous membranes close to the surface are always affected, including primarily the oral cavity (Figure 2a). Erosions predominate and can also manifest on the mucosa of the pharynx, larynx, esophagus, and genitalia (2, 3). In about half of the cases, flaccid blisters and erosions also appear on the skin, which may involve large areas. This led to a mortality of over 80% before the introduction of the corticosteroids (2, e3, 5). At present, the mortality of patients with pemphigus vulgaris is still two to three times higher than in the general population (e3, 5).

In pemphigus foliaceus, only the skin is affected, with erosions and scaly crusts, predominantly in seborrheic areas on the trunk and head (Figure 2c) (2, 3).

Paraneoplastic pemphigus is associated with neoplasia and is clinically similar to pemphigus vulgaris. Characteristic features are pronounced stomatitis, lip involvement, and polymorphic, often lichenoid, skin changes (2, 7, e17). With a mortality of 75–90%, the prognosis is unfavorable, primarily due to neoplasm and bronchiolitis obliterans, which occurs in 5–20% of cases (e3, e18, e19).

In IgA pemphigus, pustules and erosions are the most prominent lesions (e20–e22) (Table 2). Furthermore, neonatal pemphigus, pemphigus herpetiformis, and endemic pemphigus foliaceus are described as separate entities; pemphigus vegetans is considered a clinical variant of pemphigus vulgaris with predominant involvement of the axillary and in-

guinal areas. (2, 3, e23).

The differential diagnoses of pemphigus vulgaris and paraneoplastic pemphigus are severe drug reactions, such as Steven–Johnson syndrome, toxic epidermal necrolysis, stomatitis due to herpes simplex virus, hereditary epidermolysis, mucosal lichen planus, and mucous membrane pemphigoid (MMP). Pemphigus foliaceus must be differentiated from seborrheic dermatitis and impetigo, and IgA pemphigus, from pustular psoriasis as well as from pustular reactions to drugs.

Pemphigoid diseases

Bullous pemphigoid presents with tense blisters (Figure 2b), erosions, and urticarial erythema. Non-bullous forms are found in around 20% of cases (e24, e25). Characteristic features are the often severe pruritus and manifestation in old age (mean age of onset, 78 years). Therefore, bullous pemphigoid should be excluded in case of chronic pruritus in old age. Mucosal involvement can be seen in 10–20% of patients (8–10, e26).

Associated diseases that have been described include cardiovascular diseases, psoriasis, diabetes mellitus, hematological malignancies, and degenerative neurological diseases, the latter mostly preceding the skin disease and affecting 30–50% of patients (11, 12, e27–e29). Associations with the use of dipeptidyl peptidase IV inhibitors have also been observed, particularly with vildagliptin, as well as (although to a lesser degree) with spironolactone, loop diuretics, and

drugs for Parkinson's disease (13, e27, e29–e33). Gliptins should be replaced by other antidiabetic drugs in any case, and the other drugs switched to alternatives when possible. The 1-year mortality rates have been reported to range between 8% and 41% (1, e7, e10, e13, e34, e35). Differential diagnoses are bullous erysipelas, impetigo contagiosa, adverse drug reactions, herpes zoster, urticarial eczema, bullous reactions to insect stings, artifactual changes, hereditary epidermolysis, and other pemphigoid diseases.

Predominant involvement of mucous membranes supports the clinical diagnosis of MMP (Figures 2d, e). The mucous membranes of the mouth and the conjunctiva are particularly affected, as well as (less frequently) mucous membranes of the nose, pharynx, anogenital region, larynx, esophagus, and trachea. About 25–30% of patients present with additional erosions and blisters on the skin (1, 14).

Lesions of the conjunctiva, nose, larynx, esophagus and trachea in particular heal with scarring, which can lead to blindness, chronic hoarseness, difficulties in breathing and dysphagia, respectively. The main autoantigens are BP180 (in around 75% of patients) and laminin 332 (in up to 25%). Anti-laminin 332 MMP is associated with malignancy in 25–30% of cases, and in these patients, a tumor search is required (14, 15, e36). MMP has a differential diagnosis similar to that of pemphigus vulgaris.

Pemphigoid gestationis usually occurs in the third trimester of pregnancy, with severe pruritus and urticarial erythematous plaques, initially mainly in the periumbilical region. The disease resolves postpartum but usually recurs in subsequent pregnancies (1, e2, e37). As main differential diagnoses, polymorphic eruption of pregnancy and urticaria are to be distinguished. **Linear IgA** disease is characterized by tense vesicles and blisters, often arranged in an annular pattern, but may also resemble bullous pemphigoid and is a common AIBD in childhood (6, e14). In adults, induction by drugs should be considered; notably, about half of the drug-induced cases are caused by vancomycin (e38). **Anti-p200** pemphigoid clinically resembles bullous pemphigoid but shows more palmoplantar involvement (e39). In epidermolysis bullosa acquisita, the inflammatory variant mimics as bullous pemphigoid, MMP, or linear IgA disease. In the mechanobullous variant, which is present in a third of patients, blisters appear on areas most stressed by mechanical forces, such as elbows, knees, and feet. Involvement of the mucous membrane and healing with scarring are common in this variant (16, 17, e40); the most important differential diagnosis is porphyria cutanea tarda.

Dermatitis herpetiformis, which is the cutaneous manifestation of celiac disease, is characterized by severe pruritus, excoriated papules, and vesicles with predilection for knees, elbows, and buttocks (4, 18).

Diagnosis

AIBD cannot be diagnosed on the basis of the clinical

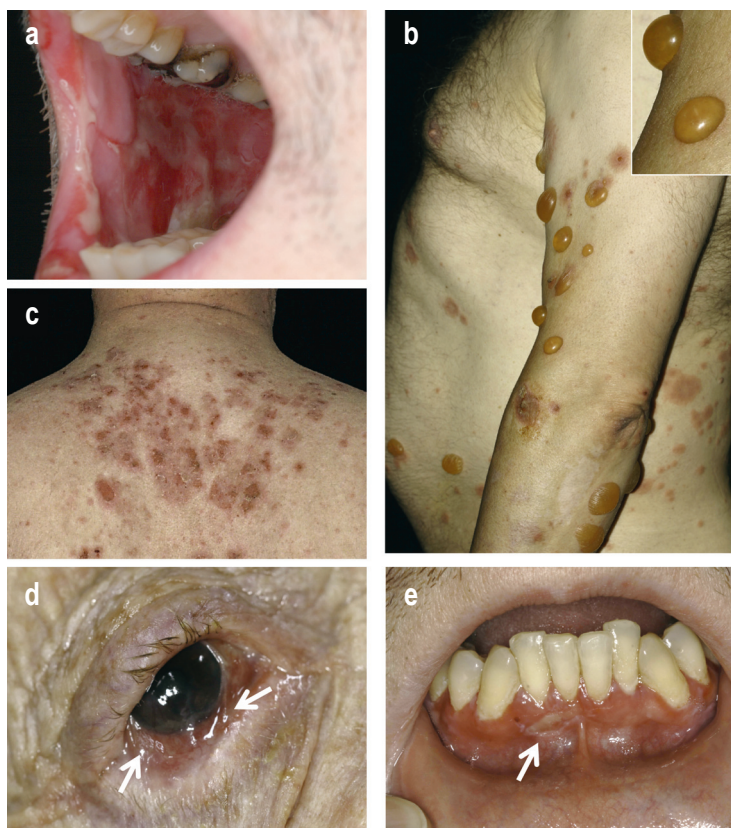


Figure 2: Clinical presentation of selected cases of autoimmune bullous dermatoses
a) Pronounced erosions of the buccal oral mucosa in pemphigus vulgaris.
b) Tense blisters, erythema, and erosions in bullous pemphigoid.
c) Erosions and crusts on the upper back in pemphigus foliaceus .
d) Conjunctival injection and synechiae (indicated by arrows).
e) Gingivitis with erosions (indicated by arrow) in MMP.

picture alone. Rather, detection of tissue-bound and/or circulating autoantibodies is required (10).

Direct immunofluorescence

Tissue-bound autoantibodies (primarily IgG and IgA) and complement deposits are detected using direct immunofluorescence (IF) in a perilesional skin/mucosal biopsy and continue to represent the gold standard in AIBD diagnostics (9, 10, 17, 18–21). Direct IF allows a differentiation between pemphigoid diseases with linear deposits on the basement membrane (Figure 3a, b), pemphigus diseases with intercellular fluorescence in the epithelium (Figure 3c), and dermatitis herpetiformis with granular deposits of IgA along the basement membrane and/or in the tips of the dermal papillae. Linear and intercellular fluorescence together indicate paraneoplastic pemphigus (7, e17).

Of the pemphigoid diseases, linear IgA disease can be differentiated based on predominant IgA deposits along the basement membrane, and epidermolysis bullosa acquisita, based on serration pattern analysis (1). Almost all pemphigoid diseases show an n-serrated pattern (Figure 3a); except for epidermolysis bullosa acquisita and bullous lupus erythematosus

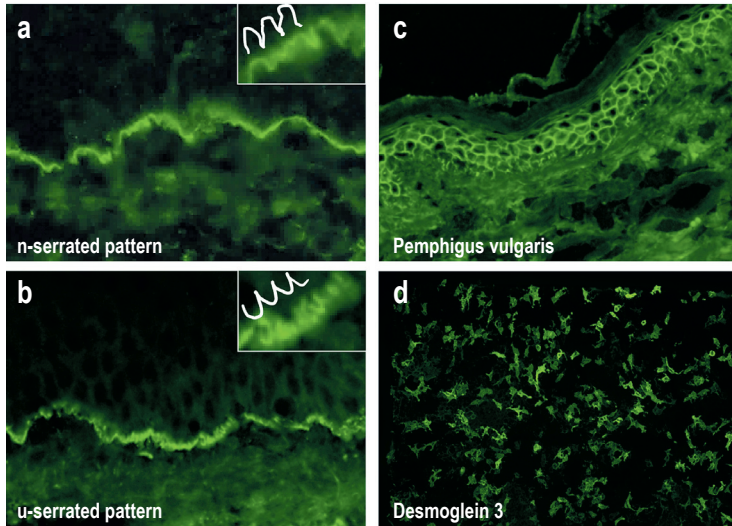


Figure 3: Direct immunofluorescence of perilesional biopsies for the detection of tissue-bound autoantibodies (a–c) and indirect immunofluorescence of the desmoglein 3-specific biochip (d). a) Linear deposits of IgG on the dermal–epidermal junction in bullous pemphigoid. At 400 x magnification, an n-serrated pattern can be seen (inset), which is found in all pemphigoid diseases with the exception of epidermolysis bullosa acquisita. b) Linear deposits of IgG on the dermal–epidermal junction zone, with a u-serrated pattern, in epidermolysis bullosa acquisita. c) Epidermal intercellular deposits of C3 with a reticular pattern, consistent with pemphigus vulgaris d) IgG reactivity to Dsg3-expressing HEK293 cells of a biochip, typical for pemphigus vulgaris

which reveal a u-serrated pattern (*Figure 3b*) (22, e41, e42).

Serological diagnostics

Circulating autoantibodies can be detected in the serum of about 90% of AIBD patients. In contrast, this is only possible for about half of the patients with epidermolysis bullosa acquisita or MMP (16, 17, e43). A serological diagnosis in combination with the clinical picture allows an exact assignment to the individual entities and thus a tailored therapy and a more precise prognosis. Anti-laminin 332 MMP and paraneoplastic pemphigus are both facultative and obligatory paraneoplasia, respectively, and a search for an underlying malignancy is indicated (7, 15, e36, e44).

To screen for suspected AIBD, an indirect IF on monkey esophagus and 1 M NaCl-split skin is carried out, which enables a differentiation into pemphigus and pemphigoid diseases (*Figure 4a–c*). The salt-split skin allows a subdivision of binding to the epidermal roof (in the case of autoantibodies against BP180 and BP230) or to the floor (autoantibodies against p200 antigen, laminin 332, and type VII collagen) of the artificial split (*Figures 1 and 4a–c, Table 2*) (1, 2, 10, 18, 19, e45).

For the detection of autoantibodies against the most important target antigens of AIBD, sensitive and specific enzyme-linked immunosorbent assays (ELISA) using the recombinant immunodominant regions of the target antigens are available (Euroim-

mun, Lübeck; MBL, Nagoya, Japan; *Table 2*) (10, e46–e52). For instance, ELISA can detect circulating antibodies against desmoglein 3 in the sera of patients with pemphigus vulgaris, and circulating antibodies against desmoglein 1 in patients with pemphigus foliaceus, in >95% of the cases (3, 23, e47, e52). Serum IgG antibodies against BP180 NC16A can be detected in 80–90% of the sera from patients with bullous pemphigoid.

The diagnostic sensitivity of bullous pemphigoid can be increased by 5–8% by the additional use of the BP230 ELISA, with which 50–60% of patients react (e46, e53). Serum autoantibodies against BP180 NC16A are also found in patients with pemphigoid gestationis as well as in 30–50% of these patients with MMP who have serum antibodies against the epidermal side of human split skin (e54–e57).

For circulating autoantibodies against desmoglein 1, desmoglein 3, BP180 NC16A, and type VII collagen, a correlation with the course of the disease has been shown (3, e47, e48, e58, e59); their determination via ELISA during the course of the disease is recommended to be included in therapy decisions (17, 21, 24). Instead of a step-by-step diagnostic approach, multivariate ELISA systems can be used in which autoantibodies against several target antigens are analyzed in parallel (e51, e60). The indirect IF-based **BIOCHIP** technology offers a comparable option. It assembles several substrates in so-called BIOCHIP mosaics in a single incubation field of a standard laboratory slide (15, 23, e61–e63) (*Figures 3d and 4d–f*).

The detection of specific autoantibodies that are not (yet) included in commercial assays is carried out in some specialized laboratories (*Table 2 and Box*).

Pathophysiology

The pathophysiology of pemphigus and pemphigoid diseases has been presented in detail in recent reviews (1–3, 25, e1, e64–e67). A common feature of all AIBDs is the presence of T cells and pathogenetically relevant autoantibodies against the respective autoantigens in genetically susceptible individuals (e68–e75). The trigger factors that lead to a breach of tolerance are still largely unknown.

In pemphigus, autoantibody binding is followed by the desmogleins being depleted from the cell surface and further signal transducing events, among others via the p38MAP kinase. Both lead to a weakening of the cell–cell interactions and to the separation of the keratinocytes/epithelial cells called acantholysis (3, 25, e1, e64).

In pemphigoid diseases, the binding of the autoantibodies leads to the local activation of complement and subsequently to the infiltration of inflammatory cells, such as eosinophils, neutrophils, macrophages, and T cells, into the upper dermis. The release of specific proteases from granulocytes, macrophages, and activated mast cells ultimately results in degradation of the proteins of the dermal–epidermal junction,

which appears histologically as subepidermal clefts and clinically as tense blisters and erosions (1, e76). C5aR1, leukotriene B4, the neonatal Fc receptor, eos-taxin, the IL-5 receptor, and IL-17A have been identified as key mediators of pemphigoid diseases; clinical studies are currently underway in which some of these are investigated (26–29, e67, e77–e83).

Therapy

German and/or European guidelines have been formulated for bullous pemphigoid, pemphigus vulgaris/foliaceus, MMP, and dermatitis herpetiformis (9, 18, 21, 24, 30, 31, e84) (eTables 1 and 2). In addition to an interdisciplinary approach with ENTs, ophthalmologists, gynecologists, general practitioners, infectiologists, paediatricians, and, if necessary, other specialist disciplines, cooperation with patient support groups is recommended, for example with the German pemphigus and pemphigoid self-help group association (*Pemphigus und Pemphigoid Selbsthilfegruppe*, www.pemphigus-pemphigoid-self-help.de) or the International Pemphigus and Pemphigoid Foundation (www.pemphigus.org).

Pemphigus diseases

First-line therapy for pemphigus vulgaris/foliaceus has changed significantly following the approval of the anti-CD20 antibody rituximab for the treatment of moderate and severe pemphigus vulgaris by the European Medicines Agency (EMA) and the US American Food and Drug Administration (FDA). Joly et al. demonstrated that treatment of patients with newly diagnosed pemphigus vulgaris/foliaceus with rituximab (2 × 1 g plus 0.5 g each, in months 12 and 18) plus prednisolone (0.5–1.0 mg/kg/day p.o. for three to six months) was significantly more effective and safer than therapy with oral prednisolone 1.0–1.5 mg/kg/day for 12–18 months (55% difference, 95% confidence interval: [38.4; 71, 7]; p < 0.0001) (32). For moderate and severe pemphigus vulgaris/foliaceus, administration of rituximab (2 × 1 g at an interval of 2–3 weeks) is recommended in combination with systemic corticosteroids (tapering over 3–6 months). Alternatively, conventional therapy with prednisolone p.o. 1.0 mg/kg/day plus azathioprine or mycophenols can be applied (eTable 2) (24). As an alternative to oral corticosteroids, intravenous corticosteroid pulses can be used (24, 31). The guideline recommends another infusion of rituximab (1 g) after six months in the event of relapse or incomplete remission; in refractory patients, the guideline also recommends high-dose intravenous immunoglobulins (IVIg) or immune apheresis (eTable 2) (24). Current clinical trials for treatment of pemphigus are evaluating efficacy and safety of inhibition of the Bruton tyrosine kinase or the neonatal Fc receptor, depletion of desmoglein 3-specific B cells using chimeric antibody receptor T cells (CAART), and tolerance induction by nanoparticles (27, 33, e1, e85, e86).

Pemphigoid diseases

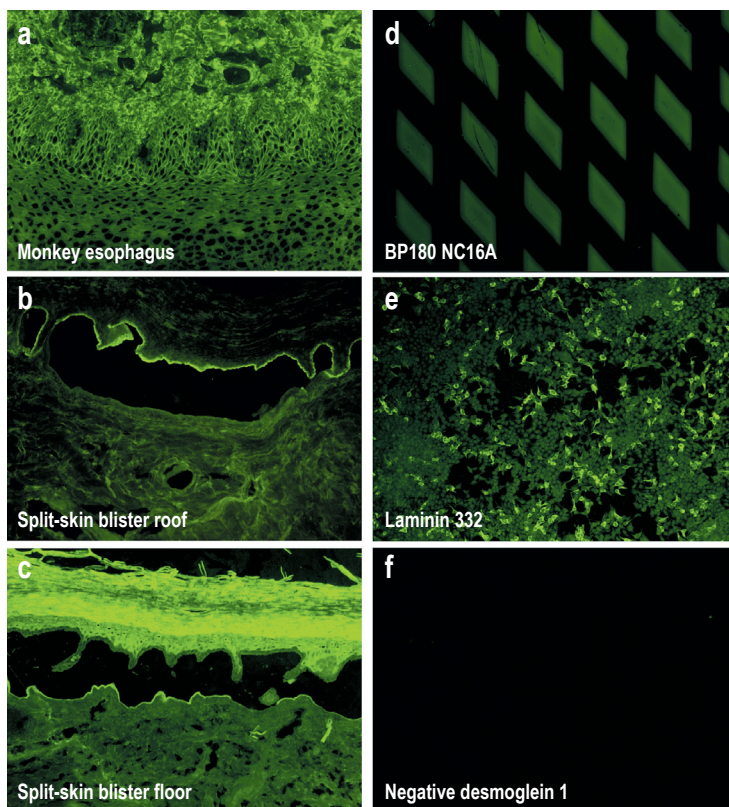


Figure 4: Indirect immunofluorescence on monkey esophagus, salt-split skin, and selected biochips

- a) IgG reactivity on monkey esophagus epithelium, with an intercellular pattern typical of pemphigus vulgaris/foliaceus
- b) IgG reactivity along the epidermal side of human salt-split skin (using 1M NaCl), fitting to binding of autoantibodies to BP180 and BP230
- c) IgG reactivity along the dermal side of human salt-split skin (using 1M NaCl), fitting to binding of autoantibodies to p200, laminin 332, and type VII collagen
- d) IgG reactivity to recombinant BP180 NC16A on a biochip, indicative of bullous pemphigoid, pemphigoid gestationis, or mucous membrane pemphigoid
- e) IgG reactivity to laminin 332-expressing HEK293 cells of a biochip, indicative of anti-laminin 332 mucous membrane pemphigoid
- f) Lack of fluorescence with Dsg1-expressing HEK293 cells of a biochip after incubation with a negative serum

TABLE 1

Incidence and Prevalence

Disease	Incidence/ million inhabitants/year	Prevalence in Germany 2014/million (e9)
Pemphigus vulgaris	0.5–6.8 (e5, 5, e88) Germany: 1.0 (e100)	94.8
Pemphigus foliaceus	< 1 (e89)	10.0
Bullous pemphigoid	6.1–42.9 (5, e5–e8, e10) Germany: 19.6 (40)	259.3
Mucous membrane pemphigoid	0.8–2 (e5, e90, e91)	24.6
Linear IgA disease	0.25–1 (e5, e92)	24.3 for children
Pemphigoid gestationis	0.08–2 (e5, e92) Germany: 2.0 (e5)	13.6 for women
Anti-p200 pemphigoid	Germany: 0.7 (40)	unknown
Epidermolysis bullosa acquisita	0.2–0.5 (5, e5–e8, e10)	2.8

TABLE 2

Target antigens of autoimmune bullous dermatoses and serological diagnostics

Disease	Target antigen	Serological diagnostics* ¹
Pemphigus diseases		
Pemphigus vulgaris	Dsg 3 Dsg 1	IIF monkey esophagus: ICF IgG+ ELISA, IIF: Dsg3+ Dsg1±
Pemphigus foliaceus	Dsg 1	IIF monkey esophagus: ICF IgG+ ELISA, IIF: Dsg1+
Paraneoplastic pemphigus	Envoplakin, periplakin, Dsg 3, desmoplakin I/II, plectin, epiplakin, BP230, BP180, Dsc 1, 2, 3, Dsg1, α2 macroglobulin-like 1	IIF monkey esophagus: ICF+, BMF± Monkey-/rat bladder: urothelium + ELISA, IIF: Dsg3±, Dsg 1±, BP180±, BP230±; ELISA: envoplakin ± IB, IP, ELISA: all other target antigens ±
IgA pemphigus	Dsc 1, 2, 3	IIF monkey esophagus: ICF IgA ELISA, IIF: Dsc1, 2, 3 IgA±; Dsg3 IgA±
Pemphigoid diseases		
Bullous pemphigoid	BP180, BP230	IIF monkey esophagus: BMF+ IIF salt-split skin: blister roof+ ELISA, IIF: BP180 (+ in 80–90%), BP230 (+ in 50–60%)
Mucous membrane pemphigoid	BP180, LAD-1, laminine332, BP230, (α4β6 integrine)* ²	IIF monkey esophagus: BMF± IIF salt-split skin: blister roof± or blister floor± ELISA: BP180±, BP230± IB: LAD-1±; BP180±; IIF: laminine332±
Linear IgA disease	LAD-1, type VII collagen	IIF monkey esophagus: BMF (IgA)+ IIF salt-split skin: blister roof (IgA) + IB, ELISA: BP180 (IgA) ±, LAD-1(IgA)+
Pemphigoid gestationis	BP180 NC16A, BP230	IIF salt-split skin: blister roof± IIF complement binding test salt-split skin: blister roof + ELISA, IIF BP180+, BP230±
Anti-p200 pemphigoid	p200 protein, laminine γ1	IIF monkey esophagus: BMF+ IIF salt-split skin: blister roof+ IB: p200+, laminine γ1±
Epidermolysis bullosa acquisita	Type VII collagen	IIF monkey esophagus: BMF± IIF salt-split skin: blister floor± ELISA, IIF: type VII collagen+
Dermatitis herpetiformis	TG2, TG3	IIF monkey esophagus (IgA): endomysium + ELISA (IgA): TG2, TG3, deamidated gliadin +

*¹ commercially available tests are indicated in bold

*² only described for individual patients

BMF, basement membrane fluorescence; Dsg, desmoglein; Dsc, desmocollin; ELISA, enzyme-linked immunosorbent assay; IIF, indirect immunofluorescence; ICF, intercellular fluorescence; IB, immunoblot; IP, immunoprecipitation; LAD-1, soluble ectodomain of BP 180; TG, transglutaminase

For bullous pemphigoid, the current German AWMF guideline recommends a whole-body application of topical 0.05% clobetasol propionate (40 g/day), a superpotent glucocorticosteroid of class IV, for mild as well as moderate cases, if necessary; for severe cases, this is usually recommended in combination with systemic treatment (24). In a controlled randomized study, topical 0.05% clobetasol propionate (40 g/day) had a comparable effect in patients with bullous pemphigoid as prednisolone (0.5 mg/kg/day) (disease control in moderate cases, topical 100% [95; 100] versus oral 95% [87; 99], p = 0.06; in severe cases, topical 99% [94; 100] versus oral 91% [83; 96], p = 0.02) (34). As a systemic treatment, prednisolone is given orally at 0.5 mg/kg/day, possibly in combination with the (poten-

tially steroid-sparing) agents azathioprine, dapsone, doxycycline, methotrexate, mycophenolate mofetil, or mycophenolate sodium. Alternatively, dapsone, doxycycline, or methotrexate can also be used as the only systemic treatment without oral corticosteroid (24) (see further details, eTable 1). In randomized controlled trials in patients with bullous pemphigoid, doxycycline was associated with significantly fewer serious adverse events than oral prednisolone (difference 19.0% [7.9; 30.1], p = 0.001), and dapsone was associated with a lower cumulative corticosteroid dose than azathioprine (p = 0.06) (35, 36). IVIg, immunoadsorption, rituximab, cyclophosphamide, or omalizumab can be used in refractory patients (eTable 1) (24, 37–39, e87).

The severity of MMP is distinguished on the basis

BOX

Dermatology departments in Germany that carry out more than 500 non-commercial test systems/year*

- Department of Dermatology, Venereology, and Allergology, University Medical Center of Schleswig-Holstein, Lübeck Campus, Lübeck, Germany
- Department of Dermatology and Allergology, University Hospital Gießen and Marburg, Marburg, Germany
- Department of Dermatology, Venereology, and Allergology and Skin Cancer Center, University Hospital Würzburg, Würzburg, Germany.

* in alphabetical order of the location (e93)

of the risk of scarring, as mild/moderate with exclusive involvement of the skin and oral mucosa, or as severe, with involvement of the eyes, nasal mucosa, pharynx, larynx, esophagus, or **trachea**. In the case of mild/moderate MMP, topical treatment with highly potent topical glucocorticoids, possibly in combination with immunomodulators, is often sufficient. For severe MMP, treatment with dapsone combined with systemic corticosteroid (prednisolone, either orally 0.5–1.5 mg/kg/day or as an intravenous pulse therapy) or cyclophosphamide (orally or intravenously) is recommended (30, e84). In the case of eye involvement, topical treatments that can be used in addition to lubricants include corticosteroids, tetracyclines, and cyclosporine (30, e84). Timely interdisciplinary treatment of inflammation is crucial before irreversible scarring occurs, especially **on the eye**.

The systemic treatments for refractory MMP and other pemphigoid diseases and dermatitis herpetiformis are summarized in *eTable 1*.

Acknowledgements

We would like to thank Carolin Mahlerwein (Lübeck) for the schematic overview figure, Ingeborg Atefi and Marina Kongsback-Reim (Lübeck) for help in preparing the fluorescent images, and the patients for the clinical images. This work was supported by the strukturelle Förderung des Exzellenz Cluster Precision Medicine in Chronic Inflammation (EXC2167) This work was supported by structural funding from the Cluster of Excellence Precision Medicine in Chronic Inflammation (PMI) (grant number EXC 2167) from the German Research Foundation.

Conflict of Interest Statement

Dr. van Beek has received reimbursement of meeting participation fees and travel expenses from Actelion, speaking honoraria from Infinite Science, and support for shared research projects from Euroimmun.

Prof. Zillikens has received consulting honoraria from Almirall, arGEN-X, Pincell, Roche Pharma, and UCB, speaking honoraria and reimbursement of travel expenses and conference fees from Novartis, Roche Pharma, Abbvie, UCB, Janssen, Almirall, and Fresenius, and support for shared research and development projects from Dompe, Euroimmun, and Fresenius.

Prof. Schmidt has received consulting honoraria from Argen X, UCB, AstraZeneca, Roche, Topas, Almirall, and Thermo Fischer, reimbursement of meeting participation fees and travel expenses as well as speaking honoraria from Biotest, Novartis, and Fresenius, and research support (third-party funds) from UCB, Biotest, Incyte, Novartis, Euroimmun, Argen X, AstraZeneca, Dompe, Admrx, Synthon/Biondis, and Fresenius.

Manuscript received on 3 August 2020, revised version accepted on 27 January 2021.

Translated from the original German by Veronica A. Raker, PhD

References

1. Schmidt E, Zillikens D: Pemphigoid diseases. *Lancet* 2013; 381: 320–32.
2. Schmidt E, Kasperkiewicz M, Joly P: Pemphigus. *Lancet* 2019; 394: 882–94.
3. Kasperkiewicz M, Ellebrecht CT, Takahashi H, et al.: Pemphigus. *Nat Rev Dis Primers* 2017; 3: 17026.
4. Sardy M, Karpati S, Merkl B, Paulsson M, Smyth N: Epidermal transglutaminase (TGase 3) is the autoantigen of dermatitis herpetiformis. *J Exp Med* 2002; 195: 747–57.
5. Langan SM, Smeeth L, Hubbard R, Fleming KM, Smith CJ, West J: Bullous pemphigoid and pemphigus vulgaris—incidence and mortality in the UK: population based cohort study. *Brmj* 2008; 337: a180.
6. Hubner F, König IR, Holtsche MM, Zillikens D, Linder R, Schmidt E: Prevalence and age distribution of pemphigus and pemphigoid diseases among pediatric patients in Germany. *J Eur Acad Dermatol Venereol* 2020; 34: 2600–5.
7. Anhalt GJ: Paraneoplastic pemphigus. *J Invest Dermatol Symp Proc* 2004; 9: 29–33.
8. Murrell DF, Daniel BS, Joly P, et al.: Definitions and outcome measures for bullous pemphigoid: recommendations by an international panel of experts. *J Am Acad Dermatol* 2012; 66: 479–85.
9. Feliciani C, Joly P, Jonkman MF, et al.: Management of bullous pemphigoid: the European Dermatology Forum consensus in collaboration with the European Academy of Dermatology and Venereology. *Br J Dermatol* 2015; 172: 867–77.
10. Schmidt E, Goebeler M, Hertl M, et al.: S2k guideline for the diagnosis of pemphigus vulgaris/foliaceus and bullous pemphigoid. *J Dtsch Dermatol Ges* 2015; 13: 713–27.
11. Kibsgaard L, Rasmussen M, Lamberg A, Deleuran M, Olesen AB, Vestergaard C: Increased frequency of multiple sclerosis among patients with bullous pemphigoid: a population-based cohort study on comorbidities anchored around the diagnosis of bullous pemphigoid. *Br J Dermatol* 2017; 176: 1486–91.
12. Schulze F, Neumann K, Recke A, Zillikens D, Linder R, Schmidt E: Malignancies in pemphigus and pemphigoid diseases. *J Invest Dermatol* 2015; 135: 1445–7.
13. Kridin K, Cohen AD: Dipeptidyl-peptidase IV inhibitor-associated bullous pemphigoid: a systematic review and meta-analysis. *J Am Acad Dermatol* 2018; S0190–9622(18)32660–4.
14. Murrell DF, Marinovic B, Caux F, et al.: Definitions and outcome measures for mucous membrane pemphigoid: recommendations of an international panel of experts. *J Am Acad Dermatol* 2015; 72: 168–74.
15. Goletz S, Probst C, Komorowski L, et al.: A sensitive and specific assay for the serological diagnosis of antilaminin 332 mucous membrane pemphigoid. *Br J Dermatol* 2019; 180: 149–56.
16. Vorobyev A, Ludwig RJ, Schmidt E: Clinical features and diagnosis of epidermolysis bullosa acquisita. *Expert Rev Clin Immunol* 2017; 13: 157–69.
17. Prost-Squarcioni C, Caux F, Schmidt E, et al.: International Bullous Diseases Group: consensus on diagnostic criteria for epidermolysis bullosa acquisita. *Br J Dermatol* 2018; 179: 30–41.
18. Görög A, Antiga E, Caproni M, et al.: S2k guideline (consensus statement) for diagnosis and therapy of dermatitis herpetiformis

- initiated by the European Academy of Dermatology and Venereology (EADV). *J Eur Acad Dermatol Venereol* 2021; 35:1251–77.
19. van Beek N, Zillikens D, Schmidt E: Diagnostik blasenbildender Autoimmundermatosen. *J Dtsch Dermatol Ges* 2018; 16: 1077–92.
 20. Harman KE, Brown D, Exton LS, et al.: British Association of Dermatologists' guidelines for the management of pemphigus vulgaris 2017. *Br J Dermatol* 2017; 177: 1170–201.
 21. Hertl M, Jedlickova H, Karpati S, et al.: Pemphigus. S2 Guideline for diagnosis and treatment—guided by the European Dermatology Forum (EDF) in cooperation with the European Academy of Dermatology and Venereology (EADV). *J Eur Acad Dermatol Venereol* 2015; 29: 405–14.
 22. Terra JB, Meijer JM, Jonkman MF, Diercks GF: The n- vs. u-serration is a learnable criterion to differentiate pemphigoid from epidermolysis bullosa acquisita in direct immunofluorescence serration pattern analysis. *Br J Dermatol* 2013; 169: 100–5.
 23. van Beek N, Kruger S, Fuhrmann T, et al.: Multicenter prospective study on multivariate diagnostics of autoimmune bullous dermatoses using the BLOCHIP(TM) technology. *J Am Acad Dermatol* 2020; 83: 1315–22.
 24. Schmidt E, Sticherling M, Sardy M, et al.: S2k guidelines for the treatment of pemphigus vulgaris/foiaceus and bullous pemphigoid: 2019 update. *J Dtsch Dermatol Ges* 2020; 18: 516–26.
 25. Spindler V, Eming R, Schmidt E, et al.: Mechanisms causing loss of keratinocyte cohesion in pemphigus. *J Invest Dermatol* 2018; 138: 32–7.
 26. Chakievskva L, Holtsche MM, Kunstner A, et al.: IL-17A is functionally relevant and a potential therapeutic target in bullous pemphigoid. *J Autoimmun* 2019; 96: 104–12.
 27. Kasprick A, Hofrichter M, Smith B, et al.: Treatment with anti-neonatal Fc receptor (FcRn) antibody ameliorates experimental epidermolysis bullosa acquisita in mice. *Br J Pharmacol* 2020; 177: 2381–92.
 28. Izumi K, Bieber K, Ludwig RJ: Current clinical trials in pemphigus and pemphigoid. *Front Immunol* 2019; 10: 978.
 29. Maglie R, Hertl M: Pharmacological advances in pemphigoid. *Curr Opin Pharmacol* 2019; 46: 34–43.
 30. Rashid H, Lambert A, Alberti-Violetti S, et al.: **European guidelines (S3) on diagnosis and management of mucous membrane pemphigoid**, initiated by the European Academy of Dermatology and Venereology (EADV) – part I: Clinical presentation and outcome measurements for disease assessment. *J Eur Acad Dermatol Venereol* (**Epub ahead of print**).
 31. Joly P, Horwath B, Patsatsi A, et al.: Updated S2K guidelines on the management of pemphigus vulgaris and foliaceus initiated by the European Academy of Dermatology and Venereology (EADV). *J Eur Acad Dermatol Venereol* 2020; 34: 1900–13.
 32. Joly P, Maho-Vaillant M, Prost-Squarcioni C, et al.: First-line rituximab combined with short-term prednisone versus prednisone alone for the treatment of pemphigus (Ritux 3): a prospective, multicentre, parallel-group, open-label randomised trial. *Lancet* 2017; 389: 2031–40.
 33. Ellebrecht CT, Bhoj VG, Nace A, et al.: Reengineering chimeric antigen receptor T cells for targeted therapy of autoimmune disease. *Science* 2016; 353: 179–84.
 34. Joly P, Roujeau JC, Benichou J, et al.: A comparison of oral and topical corticosteroids in patients with bullous pemphigoid. *N Engl J Med* 2002; 346: 321–7.
 35. Sticherling M, Franke A, Aberer E, et al.: An open, multicentre, randomized clinical study in patients with bullous pemphigoid comparing methylprednisolone and azathioprine with methylprednisolone and dapsone. *Br J Dermatol* 2017; 177: 1299–305.
 36. Williams HC, Wojnarowska F, Kirtschig G, et al.: Doxycycline versus prednisolone as an initial treatment strategy for bullous pemphigoid: a pragmatic, non-inferiority, randomised controlled trial. *Lancet* 2017; 389: 1630–8.
 37. Enk A, Hadaschik E, Eming R, et al.: European Guidelines (S1) on the use of high-dose intravenous immunoglobulin in dermatology. *J Dtsch Dermatol Ges* 2017; 15: 228–41.
 38. Kremer N, Snast I, Cohen ES, et al.: Rituximab and omalizumab for the treatment of bullous pemphigoid: a systematic review of the literature. *Am J Clin Dermatol* 2019; 20: 209–16.
 39. Ujicie H, Iwata H, Yamagami J, et al.: Japanese guidelines for the management of pemphigoid (including epidermolysis bullosa acquisita). *J Dermatol* 2019; 46: 1102–35.
 40. van Beek N, Weidinger A, Schneider SW, et al.: Incidence of pemphigoid diseases in Northern Germany in 2016—first data from the Schleswig-Holstein Registry of autoimmune bullous diseases. *J Eur Acad Dermatol Venereol* 2021; 35: 1197–202.

Corresponding author

Dr. med. Nina van Beek
 Department of Dermatology, Venereology, and Allergology
 University Medical Center Schleswig-Holstein, Campus Lübeck
 Ratzeburger Allee 160
 23538 Lübeck, Germany
 nina.vanbeek@uksh.de

Cite this as:

van Beek N, Zillikens D, Schmidt E: Bullous autoimmune dermatoses: clinical features, diagnostic evaluation, and treatment options. *Dtsch Arztebl Int* 2021; 118: 413–20. DOI: 10.3238/arztebl.m2021.0136

► **Supplementary material**

eReferences and eTables:
www.aerzteblatt-international.de/m2021.0136

Questions on the article in the issue 24/2021:

Bullous Autoimmune Dermatoses – Clinical Features, Diagnostic Evaluation, and Treatment Options

CME credit for this unit can be obtained via cme.aerzteblatt.de until 17 June 2022.

Only one answer is possible per question. Please select the answer that is most appropriate.

Question 1

Which type of autoimmune bullous diseases mainly has antibodies targeted against the desmoglein 3 antigen?

- a) IgA pemphigus
- b) Bullous pemphigoid
- c) Linear IgA dermatosis
- d) Pemphigus vulgaris
- e) Mucous membrane pemphigoid (MMP)

Question 2

Which autoimmune bullous disease has the highest prevalence in Germany?

- a) Pemphigus vulgaris
- b) Pemphigoid gestationis
- c) Bullous pemphigoid
- d) Epidermolysis bullosa acquisita
- e) Pemphigus foliaceus

Question 3

Which is characteristic of bullous pemphigoid?

- a) Scaly crusts, manifestation in middle age
- b) Manifestation mostly in childhood, severe pruritus
- c) Erosion strictly limited to the skin, severe pruritus
- d) Manifestation at a young age, usually without pruritus
- e) Manifestation in old age, severe pruritus

Question 4

Which autoimmune bullous dermatosis almost always occurs in connection with celiac disease?

- a) Epidermolysis bullosa acquisita
- b) Dermatitis herpetiformis
- c) Linear IgA disease
- d) Anti-p200 pemphigoid
- e) IgA pemphigus

Question 5

Detection of autoantibodies is part of the serological diagnosis of patients with autoimmune bullous diseases. In what percentage of these patients can circulating autoantibodies be detected in the serum?

- a) ca. 90 %
- b) ca. 70 %
- c) ca. 50 %
- d) ca. 20 %
- e) ca. 10 %

Question 6

Which technology is used to detect autoantibodies against the most important target antigens in autoimmune bullous dermatoses?

- a) Polymerase chain reaction (PCR)
- b) Immunoprecipitation
- c) Fluorescence In Situ Hybridization (FISH)
- d) Matrix-Assisted Laser-Desorption-Ionization (MALDI)
- e) Enzyme-linked immunosorbent assay (ELISA)

Question 7

In addition to conventional therapy, which treatment is recommended for moderate and severe pemphigus vulgaris?

- a) Rituximab (1 g/day for 3 months) as a monotherapy
- b) Azathioprine as monotherapy
- c) Rituximab in combination with systemic corticosteroids
- d) Mycophenolate mofetil (500 mg/day) as a monotherapy
- e) Azathioprine combined with mycophenolate mofetil

Question 8

Which types of autoimmune bullous diseases are most common in children?

- a) Linear IgA dermatosis and pemphigus vulgaris
- b) Bullous pemphigoid and mucous membrane pemphigoid (MMP)
- c) Epidermolysis bullosa acquisita and pemphigus foliaceus
- d) IgA pemphigus and dermatitis herpetiformis Dühring
- e) Dermatitis herpetiformis Dühring and bullous pemphigoid

Question 9

Which pattern is typically seen in the direct immunofluorescence of perilesional biopsies in epidermolysis bullosa acquisita?

- a) An N-serrated pattern
- b) A U-serrated pattern
- c) An A-serrated pattern
- d) An O-serrated pattern
- e) A Y-serrated pattern

Question 10

What is known about the occurrence of pemphigoid gestationis?

- a) It is usually most pronounced at the beginning of pregnancy
- b) The disease stops immediately postpartum
- c) After recovery from the disease, it will not occur in subsequent pregnancies
- d) Urticarial erythematous plaques do not appear on the abdomen
- e) It usually occurs in the third trimester of pregnancy

Supplementary material:

Bullous Autoimmune Dermatoses

Clinical Features, Diagnostic Evaluation, and Treatment Options

by Nina van Beek, Dettlef Zillikens, and Enno Schmidt

Dtsch Arztebl Int 2021; 118: 413–20. DOI: 10.3238/arztebl.m2021.0136

eReferences

e1. Pollmann R, Schmidt T, Eming R, Hertl M: Pemphigus: a comprehensive review on pathogenesis, clinical presentation and novel therapeutic approaches. *Clin Rev Allergy Immunol* 2018; 54: 1–25.

e2. Amber KT, Murrell DF, Schmidt E, Joly P, Borradori L: Autoimmune subepidermal bullous diseases of the skin and mucosae: clinical features, diagnosis, and management. *Clin Rev Allergy Immunol* 2018; 54: 26–51.

e3. Kridin K: Pemphigus group: overview, epidemiology, mortality, and comorbidities. *Immunol Res* 2018; 66: 255–70.

e4. Kridin K, Ludwig RJ: The growing incidence of bullous pemphigoid: overview and potential explanations. *Front Med (Lausanne)* 2018; 5: 220.

e5. Bertram F, Brocker EB, Zillikens D, Schmidt E: Prospective analysis of the incidence of autoimmune bullous disorders in Lower Franconia, Germany. *J Dtsch Dermatol Ges* 2009; 7: 434–40.

e6. Marazza G, Pham HC, Scharer L, et al.: Incidence of bullous pemphigoid and pemphigus in Switzerland: a 2-year prospective study. *Br J Dermatol* 2009; 161: 861–8.

e7. Joly P, Baricault S, Sparsa A, et al.: Incidence and mortality of bullous pemphigoid in France. *J Invest Dermatol* 2012; 132: 1998–2004.

e8. Forsti AK, Jokelainen J, Timonen M, Tasanen K: Increasing incidence of bullous pemphigoid in Northern Finland: a retrospective database study in Oulu University Hospital. *Br J Dermatol* 2014; 171: 1223–6.

e9. Hubner F, Recke A, Zillikens D, Linder R, Schmidt E: Prevalence and age distribution of pemphigus and pemphigoid diseases in Germany. *J Invest Dermatol* 2016; 136: 2495–8.

e10. Jelti L, Cordel N, Gillibert A, et al.: Incidence and mortality of pemphigus in France. *J Invest Dermatol* 2019; 139: 469–73.

e11. Joly P: Incidence of bullous pemphigoid and pemphigus vulgaris. *BMJ* 2008; 337: a209.

e12. Loget J, Barbe C, Duvert-Lehembre S, et al.: The regibul register: a tool for monitoring the distribution and incidence of autoimmune bullous dermatoses in three french regions, 2010 to 2015. *Acta Derm Venereol* 2018; 98: 380–1.

e13. Persson MSM, Harman KE, Vinogradova Y, et al.: Incidence, prevalence and mortality of bullous pemphigoid in England 1998–2017: a population-based cohort study. *Br J Dermatol* 2021; 184: 68–77.

e14. van Beek N, Schmidt E: Autoimmune bullous diseases. In: Höger P, Kinsler V, Yan A (Eds) *Harper's Textbook of Pediatric Dermatology*, chapter 73, 4th edition Wiley-Blackwell, Chichester 2020: 868–97.

e15. Sarig O, Bercovici S, Zoller L, et al.: Population-specific association between a polymorphic variant in ST18, encoding a pro-apoptotic molecule, and pemphigus vulgaris. *J Invest Dermatol* 2012; 132: 1798–805.

e16. Hirose M, Schilf P, Benoit S, et al.: Polymorphisms in the mitochondrially encoded ATP synthase 8 gene are associated with susceptibility to bullous pemphigoid in the German population. *Exp Dermatol* 2015; 24: 715–7.

e17. Zimmermann J, Bahmer F, Rose C, Zillikens D, Schmidt E: Clinical and immunopathological spectrum of paraneoplastic pemphigus. *J Dtsch Dermatol Ges* 2010; 8: 598–606.

e18. Yong AA, Tey HL: Paraneoplastic pemphigus. *Australas J Dermatol* 2013; 54: 241–50.

e19. Leger S, Picard D, Ingen-Housz-Oro S, et al.: Prognostic factors of paraneoplastic pemphigus. *Arch Dermatol* 2012; 148: 1165–72.

e20. Hashimoto T, Kiyokawa C, Mori O, et al.: Human desmocollin 1 (Dsc1) is an autoantigen for the subcorneal pustular dermatosis type of IgA pemphigus. *J Invest Dermatol* 1997; 109: 127–31.

e21. Muller R, Heber B, Hashimoto T, et al.: Autoantibodies against desmocollins in European patients with pemphigus. *Clin Exp Dermatol* 2009; 34: 898–903.

e22. Hashimoto T, Teye K, Ishii N: Clinical and immunological studies of 49 cases of various types of intercellular IgA dermatosis and 13 cases of classical subcorneal pustular dermatosis examined at Kurume University. *Br J Dermatol* 2017; 176: 168–75.

e23. Kasperkiewicz M, Kowalewski C, Jablonska S: Pemphigus herpeticiformis: from first description until now. *J Am Acad Dermatol* 2014; 70: 780–7.

e24. della Torre R, Combescurie C, Cortes B, et al.: Clinical presentation and diagnostic delay in bullous pemphigoid: a prospective nationwide cohort. *Br J Dermatol* 2012; 167: 1111–7.

e25. Lamberts A, Meijer JM, Jonkman MF: Nonbullous pemphigoid: a systematic review. *J Am Acad Dermatol* 2018; 78: 989–95 e2.

e26. Kridin K, Bergman R: Assessment of the prevalence of mucosal involvement in bullous pemphigoid. *JAMA Dermatol* 2019; 155: 166–71.

e27. Forsti AK, Huilaja L, Schmidt E, Tasanen K: Neurological and psychiatric associations in bullous pemphigoid—more than skin deep? *Exp Dermatol* 2017; 26: 1228–34.

e28. Bech R, Kibsgaard L, Vestergaard C: Comorbidities and treatment strategies in bullous pemphigoid: an appraisal of the existing literature. *Front Med (Lausanne)* 2018; 5: 238.

e29. Bastuji-Garin S, Joly P, Lemordant P, et al.: Risk factors for bullous pemphigoid in the elderly: a prospective case-control study. *J Invest Dermatol* 2011; 131: 637–43.

e30. Lloyd-Lavery A, Chi CC, Wojnarowska F, Taghipour K: The associations between bullous pemphigoid and drug use: a UK case-control study. *JAMA Dermatol* 2013; 149: 58–62.

e31. Varpuluoma O, Forsti AK, Jokelainen J, et al.: Vildagliptin significantly increases the risk of bullous pemphigoid: a finnish nationwide registry study. *J Invest Dermatol* 2018; 138: 1659–61.

e32. Plaquet M, Tetart F, Fardet L, et al.: Higher frequency of dipeptidyl peptidase-4 inhibitor intake in bullous pemphigoid patients than in the french general population. *J Invest Dermatol* 2019; 139: 835–41.

e33. Liu SD, Chen WT, Chi CC: Association between medication use and bullous pemphigoid: a systematic review and meta-analysis. *JAMA Dermatol* 2020; 156: 891–900.

e34. Holtsche MM, Goletz S, van Beek N, et al.: Prospective study in bullous pemphigoid: association of high serum anti-BP180 IgG levels with increased mortality and reduced Karnofsky score. *Br J Dermatol* 2018; 179: 918–24.

e35. Kridin K, Shihade W, Bergman R: Mortality in patients with bullous pemphigoid: a retrospective cohort study, systematic review and meta-analysis. *Acta Derm Venereol* 2019; 99: 72–7.

e36. Egan CA, Lazarova Z, Darling TN, Yee C, Cote T, Yancey KB: Anti-epiligrin cicatricial pemphigoid and relative risk for cancer. *Lancet* 2001; 357: 1850–1.

e37. Huilaja L, Makikallio K, Tasanen K: Gestational pemphigoid. *Orphanet J Rare Dis* 2014; 9: 136.

e38. Lammer J, Hein R, Roenneberg S, Biedermann T, Volz T: Drug-induced linear IgA bullous dermatosis: a case report and review of the literature. *Acta Derm Venereol* 2019; 99: 508–15.

e39. Goletz S, Hashimoto T, Zillikens D, Schmidt E: Anti-p200 pemphigoid. *J Am Acad Dermatol* 2014; 71: 185–91.

e40. Ludwig RJ: Clinical presentation, pathogenesis, diagnosis, and treatment of epidermolysis bullosa acquisita. *ISRN dermatology* 2013; 2013: 812029.

e41. Meijer JM, Atefi I, Diercks GFH, et al.: Serration pattern analysis for differentiating epidermolysis bullosa acquisita from other pemphigoid diseases. *J Am Acad Dermatol* 2018; 78: 754–59 e6.

e42. Buijsrogge JJ, Diercks GF, Pas HH, Jonkman MF: The many faces of epidermolysis bullosa acquisita after serration pattern analysis by direct immunofluorescence microscopy. *Br J Dermatol* 2011; 165: 92–8.

- e43. Holtsche MM, Zillikens D, Schmidt E: [Mucous membrane pemphigoid]. *Hautarzt* 2018; 69: 67–83.
- e44. Kaplan I, Hodak E, Ackerman L, Mimouni D, Anhalt GJ, Calderon S: Neoplasms associated with paraneoplastic pemphigus: a review with emphasis on non-hematologic malignancy and oral mucosal manifestations. *Oral Oncol* 2004; 40: 553–62.
- e45. Sardy M, Kostaki D, Varga R, Peris K, Ruzicka T: Comparative study of direct and indirect immunofluorescence and of bullous pemphigoid 180 and 230 enzyme-linked immunosorbent assays for diagnosis of bullous pemphigoid. *J Am Acad Dermatol* 2013; 69: 748–53.
- e46. Blocker IM, Dahnrich C, Probst C, et al.: Epitope mapping of BP230 leading to a novel enzyme-linked immunosorbent assay for autoantibodies in bullous pemphigoid. *Br J Dermatol* 2012; 166: 964–70.
- e47. Schmidt E, Dahnrich C, Rosemann A, et al.: Novel ELISA systems for antibodies to desmoglein 1 and 3: correlation of disease activity with serum autoantibody levels in individual pemphigus patients. *Exp Dermatol* 2010; 19: 458–63.
- e48. Kobayashi M, Amagai M, Kuroda-Kinoshita K, et al.: BP180 ELISA using bacterial recombinant NC16a protein as a diagnostic and monitoring tool for bullous pemphigoid. *J Dermatol Sci* 2002; 30: 224–32.
- e49. Saleh MA, Ishii K, Kim YJ, et al.: Development of NC1 and NC2 domains of type VII collagen ELISA for the diagnosis and analysis of the time course of epidermolysis bullosa acquisita patients. *J Dermatol Sci* 2011; 62: 169–75.
- e50. Sitaru C, Dahnrich C, Probst C, et al.: Enzyme-linked immunosorbent assay using multimers of the 16th non-collagenous domain of the BP180 antigen for sensitive and specific detection of pemphigoid autoantibodies. *Exp Dermatol* 2007; 16: 770–7.
- e51. van Beek N, Dahnrich C, Johannsen N, et al.: Prospective studies on the routine use of a novel multivalent enzyme-linked immunosorbent assay for the diagnosis of autoimmune bullous diseases. *J Am Acad Dermatol* 2017; 76: 889–94. e5.
- e52. Ishii K, Amagai M, Hall RP, et al.: Characterization of autoantibodies in pemphigus using antigen-specific enzyme-linked immunosorbent assays with baculovirus-expressed recombinant desmogleins. *J Immunol* 1997; 159: 2010–7.
- e53. Charneux J, Lorin J, Vitry F, et al.: Usefulness of BP230 and BP180-NC16a enzyme-linked immunosorbent assays in the initial diagnosis of bullous pemphigoid: a retrospective study of 138 patients. *Arch Dermatol* 2011; 147: 286–91.
- e54. Sadik CD, Pas HH, Bohlmann MK, et al.: Value of BIOCHIP technology in the serological diagnosis of pemphigoid gestationis. *Acta Derm Venereol* 2017; 97: 128–30.
- e55. Murakami H, Nishioka S, Setterfield J, et al.: Analysis of antigens targeted by circulating IgG and IgA autoantibodies in 50 patients with cicatricial pemphigoid. *J Dermatol Sci* 1998; 17: 39–44.
- e56. Calabresi V, Carozzo M, Cozzani E, et al.: Oral pemphigoid autoantibodies preferentially target BP180 ectodomain. *Clin Immunol* 2007; 122: 207–13.
- e57. Schmidt E, Skrobek C, Kromminga A, et al.: Cicatricial pemphigoid: IgA and IgG autoantibodies target epitopes on both intra- and extracellular domains of bullous pemphigoid antigen 180. *Br J Dermatol* 2001; 145: 778–83.
- e58. Schmidt E, Obe K, Bocker EB, Zillikens D: Serum levels of autoantibodies to BP180 correlate with disease activity in patients with bullous pemphigoid. *Arch Dermatol* 2000; 136: 174–8.
- e59. Kim JH, Kim YH, Kim S, et al.: Serum levels of anti-type VII collagen antibodies detected by enzyme-linked immunosorbent assay in patients with epidermolysis bullosa acquisita are correlated with the severity of skin lesions. *J Eur Acad Dermatol Venereol* 2012; 27: e224–30.
- e60. Horvath ON, Varga R, Kaneda M, Schmidt E, Ruzicka T, Sardy M: Diagnostic performance of the „MESACUP anti-Skin profile TEST“. *Eur J Dermatol* 2016; 26: 56–63.
- e61. van Beek N, Rentzsch K, Probst C, et al.: Serological diagnosis of autoimmune bullous skin diseases: prospective comparison of the BIOCHIP mosaic-based indirect immunofluorescence technique with the conventional multi-step single test strategy. *Orphanet J Rare Dis* 2012; 7: 49.
- e62. Yang A, Xuan R, Melbourne W, Tran K, Murrell DF: Validation of the BIOCHIP test for the diagnosis of bullous pemphigoid, pemphigus vulgaris and pemphigus foliaceus. *J Eur Acad Dermatol Venereol* 2020; 34: 153–60.
- e63. Komorowski L, Muller R, Vorobyev A, et al.: Sensitive and specific assays for routine serological diagnosis of epidermolysis bullosa acquisita. *J Am Acad Dermatol* 2012; 68: e89–95.
- e64. Waschke J, Spindler V: Desmosomes and extradesmosomal adhesive signaling contacts in pemphigus. *Med Res Rev* 2014; 34: 1127–45.
- e65. Hammers CM, Stanley JR: Mechanisms of disease: pemphigus and bullous pemphigoid. *Annu Rev Pathol* 2016; 11: 175–97.
- e66. Sadik CD, Schmidt E, Zillikens D, Hashimoto T: Recent progresses and perspectives in autoimmune bullous diseases. *J Allergy Clin Immunol* 2020; 145: 1145–7.
- e67. Sadik CD, Zillikens D: Current treatments and developments in pemphigoid diseases as paradigm diseases for antibody-driven, organ-specific autoimmune diseases. *Semin Hematol* 2016; 53 (Suppl 1): S51–3.
- e68. Anhalt GJ, Labib RS, Voorhees JJ, Beals TF, Diaz LA: Induction of pemphigus in neonatal mice by passive transfer of IgG from patients with the disease. *N Engl J Med* 1982; 306: 1189–96.
- e69. Amagai M, Tsunoda K, Suzuki H, Nishifuji K, Koyasu S, Nishikawa T: Use of autoantigen-knockout mice in developing an active autoimmune disease model for pemphigus. *J Clin Invest* 2000; 105: 625–31.
- e70. Eming R, Hennerici T, Backlund J, et al.: Pathogenic IgG antibodies against desmoglein 3 in pemphigus vulgaris are regulated by HLA-DRB1*04:02-restricted T cells. *J Immunol* 2014; 193: 4391–9.
- e71. Liu Z, Diaz LA, Troy JL, et al.: A passive transfer model of the organ-specific autoimmune disease, bullous pemphigoid, using antibodies generated against the hemidesmosomal antigen, BP180. *J Clin Invest* 1993; 92: 2480–8.
- e72. Nishie W, Sawamura D, Goto M, et al.: Humanization of autoantigen. *Nat Med* 2007; 13: 378–83.
- e73. Sitaru C, Mihai S, Otto C, et al.: Induction of dermal-epidermal separation in mice by passive transfer of antibodies specific to type VII collagen. *J Clin Invest* 2005; 115: 870–8.
- e74. Haeberle S, Wei X, Bieber K, et al.: Regulatory T-cell deficiency leads to pathogenic bullous pemphigoid antigen 230 autoantibody and autoimmune bullous disease. *J Allergy Clin Immunol* 2018; 142: 1831–42. e7.
- e75. Heppel EN, Tofern S, Schulze FS, et al.: Experimental laminin 332 mucous membrane pemphigoid critically involves C5aR1 and reflects clinical and immunopathological characteristics of the human disease. *J Invest Dermatol* 2017; 137: 1709–18.
- e76. Sadik CD, Schmidt E: Resolution in bullous pemphigoid. *Semin Immunopathol* 2019; 41: 645–54.
- e77. Karsten CM, Pandey MK, Figge J, et al.: Anti-inflammatory activity of IgG1 mediated by Fc galactosylation and association of FcγRIIB and dectin-1. *Nat Med* 2012; 18: 1401–6.
- e78. Koga H, Kasprick A, Lopez R, et al.: Therapeutic effect of a novel phosphatidylinositol-3-kinase delta inhibitor in experimental epidermolysis bullosa acquisita. *Front Immunol* 2018; 9: 1558.
- e79. Samavedam UK, Mitschker N, Kasprick A, et al.: Whole-genome expression profiling in skin reveals SYK as a key regulator of inflammation in experimental epidermolysis bullosa acquisita. *Front Immunol* 2018; 9: 249.
- e80. Stussel P, Dieckhoff KS, Kunzel S, et al.: Propranolol is an effective topical and systemic treatment option for experimental epidermolysis bullosa acquisita. *J Invest Dermatol* 2020; 140: 2408–20.
- e81. Gunther C, Wozel G, Meurer M, Pfeiffer C: Up-regulation of CCL11 and CCL26 is associated with activated eosinophils in bullous pemphigoid. *Clin Exp Immunol* 2011; 166: 145–53.
- e82. Shrikhande M, Hunziker T, Braathen LR, Pichler WJ, Dahinden CA, Yawalkar N: Increased coexpression of eotaxin and interleukin 5 in bullous pemphigoid. *Acta Derm Venereol* 2000; 80: 277–80.
- e83. Wakugawa M, Nakamura K, Hino H, et al.: Elevated levels of eotaxin and interleukin-5 in blister fluid of bullous pemphigoid: correlation with tissue eosinophilia. *Br J Dermatol* 2000; 143: 112–6.
- e84. Schmidt E, Lambert A, Marzano A, et al.: S3 guideline on the diagnosis and management of mucous membrane pemphigoid initiated by the European Academy of Dermatology and Venereology (EADV). *J Eur Acad Dermatol Venereol* (in preparation).
- e85. Lee A, Sandhu S, Imlay-Gillespie L, Mulligan S, Shumack S: Successful use of Bruton's kinase inhibitor, ibrutinib, to control paraneoplastic pemphigus in a patient with paraneoplastic autoimmune multiorgan syndrome and chronic lymphocytic leukaemia. *Australas J Dermatol* 2017; 58: e240–e2.
- e86. Hofrichter M, Dworschak J, Emtenani S, et al.: Immunoadsorption of desmoglein-3-specific IgG abolishes the blister-inducing capacity of pemphigus vulgaris IgG in neonatal mice. *Front Immunol* 2018; 9: 1935.

- e87. Hubner F, Kasperkiewicz M, Knuth-Rehr D, et al.: Adjuvant treatment of severe/refractory bullous pemphigoid with protein A immunoadsorption. *J Dtsch Dermatol Ges* 2018; 16: 1109–18.
- e88. Bastuji-Garin S, Souissi R, Blum L, et al.: Comparative epidemiology of pemphigus in Tunisia and France: unusual incidence of pemphigus foliaceus in young Tunisian women. *J Invest Dermatol* 1995; 104: 302–5.
- e89. Stanley J: Pemphigus. In: Wolff K GL, Katz SI, et al. (ed.): *Fitzpatrick's dermatology in general medicine*. New York: McGraw-Hill 2008; p. p. 459–68.
- e90. Bernard P, Vaillant L, Labeille B, et al.: Incidence and distribution of subepidermal autoimmune bullous skin diseases in three French regions. Bullous Diseases French Study Group. *Arch Dermatol* 1995; 131: 48–52.
- e91. Radford CF, Rauz S, Williams GP, Saw VP, Dart JK: Incidence, presenting features, and diagnosis of cicatrizing conjunctivitis in the United Kingdom. *Eye (Lond)* 2012; 26: 1199–208.
- e92. Milinkovic MV, Jankovic S, Medenica L, et al.: Incidence of autoimmune bullous diseases in Serbia: a 20-year retrospective study. *J Dtsch Dermatol Ges* 2016; 14: 995–1005.
- e93. van Beek N, Knuth-Rehr D, Altmeyer P, et al.: Diagnostics of autoimmune bullous diseases in German dermatology departments. *J Dtsch Dermatol Ges* 2012; 10: 492–9.
- e94. Tsuruta D, Ishii N, Hamada T, et al.: IgA pemphigus. *Clin Dermatol* 2011; 29: 437–42.
- e95. Kridin K, Patel PM, Jones VA, Cordova A, Amber KT: IgA pemphigus: a systematic review. *J Am Acad Dermatol* 2020; 82: 1386–92.
- e96. Wojnarowska F, Kirtschig G, Khumalo N: Treatment of subepidermal immunobullous diseases. *Clin Dermatol* 2001; 19: 768–77.
- e97. Kasperkiewicz M, Meier M, Zillikens D, Schmidt E: Linear IgA disease: successful application of immunoadsorption and review of the literature. *Dermatology* 2010; 220: 259–63.
- e98. Iwata H, Vorobyev A, Koga H, et al.: Meta-analysis of the clinical and immunopathological characteristics and treatment outcomes in epidermolysis bullosa acquisita patients. *Orphanet J Rare Dis* 2018; 13: 153.
- e99. Santi CG, Gripp AC, Roselino AM, et al.: Consensus on the treatment of autoimmune bullous dermatoses: bullous pemphigoid, mucous membrane pemphigoid and epidermolysis bullosa acquisita—Brazilian Society of Dermatology. *An Bras Dermatol* 2019; 94: 33–47.
- e100. Hahn-Ristic K, Rzany B, Amagai M, Bröcker E-B, Zillikens D: Increased incidence of pemphigus vulgaris in southern Europeans living in Germany compared with native Germans. *Eur Acad Dermatol Venereol* 2002; 16:68–71.

eTABLE 1

Treatment options for pemphigoid diseases and dermatitis herpetiformis

Disease/severity	Treatment	
	Medication ^{a,2}	Adverse drug reactions ^{a,11}
Bullous pemphigoid^{a,1} (24)		
Mild (< 10% body surface area)	Topical 0.05% clobetasol propionate (level B)	Skin atrophy
Moderate (10–30% body surface area)	Topical 0.05% clobetasol propionate (level B) ± systemic treatment Systemic treatment Prednisolone ^{a,3} p.o. 0.5 mg/kg/day (level B) or Dapsone ^{a,7} ± prednisolone ^{a,3} p.o. 0.5 mg/kg/day (level C) or Doxycycline (200 mg/day) (level B) ± nicotinamide (level D) ± prednisolone ^{a,3} p.o. 0.5 mg/kg/day or Azathioprine ^{a,5} (level C) + prednisolone ^{a,3} p.o. 0.5 mg/kg/day or Methotrexate (≤ 20 mg /week) (level D) ± prednisolone ^{a,3} p.o. 0.5 mg/kg/day or Mycophenolis ^{a,6} (level C) + prednisolone ^{a,3} p.o. 0.5 mg/kg/day	Skin atrophy Hypertension, diabetes, osteoporosis, AI, infections Dapsone: hemolytic anemia, GI complaints, methemoglobinemia, agranulocytosis Doxycycline: GI complaints, sensitivity to light Azathioprine: myelo- and hepatotoxicity, arthralgias, infections Methotrexate: myelo- and hepatotoxicity, infections, kidney dysfunction, possible reactivation Mycophenolis: GI complaints, infections, (rarely) myelotoxicity (see above)
Severe (> 30% body surface area)	Topical 0.05% clobetasol propionate + systemic treatment (see above)	(see above)
Refractory to treatment	IVIg (level D) ^{a,9} Immunoadsorption (level D) ^{a,10} Rituximab ^{a,8} (level D) Cyclophosphamide (level E) Omalizumab (level E)	Headache, chills, flushing, fever, hypertension, nausea Hypotension, thrombosis, bradycardia, anaphylaxis, herpes zoster infection, sepsis Infusion reaction, infections, reactivation of hepatitis B, PML Myelotoxicity, hemorrhagic cystitis, carcinogenicity, infertility, infections

Disease/ severity	Treatment	
	Medication ^{*2}	Adverse drug reactions ^{*11}
MMP^{*1} (30, e84)		
Mild/moderate (only oral cavity and skin are affected)	Topical corticosteroids class III/IV and/or dapsone ^{*7} (level D), methotrexate (≤ 20 mg/week) (level E) or tetracycline (level E)	(ADRs of these drugs are described above) GI complaints, dizziness
Mild, refractory to treatment	+ Prednisolone ^{*3} p.o. 0.5–1.5 mg/kg/day (level C) and/or azathioprine ^{*5} or mycophenolis ^{*6} (level D)	(ADRs of these drugs are described above)
Severe (involvement of conjunctiva, nasopharynx, larynx, trachea or esophagus)	Dapsone ^{*7} (level C) + cyclophosphamide p.o./i.v. and/or prednisolone ^{*3} p.o. 0.5–1.5 mg/kg/day (or i.v. steroid pulse ^{*4}) (all: level D)	(ADRs of these drugs are described above)
Severe, refractory to treatment	Dapsone ^{*7} + rituximab 2 × 1 g ^{*8} (level D) if still refractory; + IVIg ^{*9} (level D)	(ADRs of these drugs are described above)
Linear IgA disease (e96, e97)		
Refractory to treatment	Topical corticosteroids class III/IV or prednisolone ^{*3} p.o. 0.25–0.5 mg/kg/day ± dapsone ^{*7} (all: level D)	(ADRs of these drugs are described above)
	Prednisolone ^{*3} p.o. 0.5 mg/kg/day + Sulfasalazine/pyridine (level D)	(see above) GI complaints, hepato- and nephrotoxicity, myelotoxicity
	Doxycycline (200 mg/day) (level E)	(see above)
	Cholchicine (level E)	GI complaints, hepato- and nephrotoxicity, myelotoxicity
	IVIg ^{*9} , azathioprine ^{*5} , or mycophenolis ^{*6} (all, level E)	(see above)
Pemphigoid gestationis (e37)	Topical 0.05% clobetasol propionate or prednisolone ^{*3} p.o. 0.25–0.5 mg/kg/day (level D)	(ADRs of these drugs are described above)
Refractory to treatment	Immunoadsorption ^{*10} , rituximab ^{*6} (only postpartum) (level E)	(ADRs of these drugs are described above)
Anti-p200 pemphigoid (e39)	Topical 0.05% clobetasol propionate ± prednisolone ^{*3} p.o. 0.25–0.5 mg/kg/day ± dapsone ^{*7} or doxycycline 200 mg/day (level E)	(ADRs of these drugs are described above)
Epidermolysis bullosa acquisita (39, e98, e99)		
Mild (< 10% body surface area)	Topical 0.05% clobetasol propionate + dapsone ^{*7} or colchicine (level E)	(ADRs of these drugs are described above)
Moderate/severe	Prednisolone ^{*3} p.o. 1.0–2.0 mg/kg/day or i.v. steroid pulses ^{*4} , tapering over course+ azathioprine ^{*5} , mycophenolis ^{*6} or dapsone ^{*7} (all: level E)	(ADRs of these drugs are described above)
Refractory to treatment	+ Rituximab ^{*6} and/or IVIg ^{*9} (both level E)	(ADRs of these drugs are described above)
Dermatitis herpetiformis^{*1} (18)	Gluten-free diet (life-long) ± dapsone ^{*7} (until skin lesions have healed) (level D)	(ADRs of these drugs are described above)

*1 German and/or European therapy guidelines are available for these diseases

*2 Level of evidence: level A, meta-analyses of prospective, controlled trials; level B, high-quality prospective, controlled trials; level C, lower-quality prospective, controlled trials; level D, larger case studies; level E, small case studies, case reports

*3 or prednisolone equivalent

*4 dexamethasone 100 mg/day for three consecutive days, or prednisolone 500–1000 mg/day, every 3–4 weeks, eventually every 6–8 weeks

*5 2.0–2.5 mg/kg/day with normal thiopurine methyltransferase levels

*6 mycophenolate mofetil (2 g/day), mycophenolate sodium (1440 mg/day)

*7 1.0–1.5 mg/kg/day with normal glucose 6-phosphate levels

*8 or biosimilar

*9 2 g/kg for 2–5 days every four weeks; after six months, interval extension (37)

*10 on 3–4 consecutive days with regenerative adsorbers every 3–4 weeks, for 8–16 weeks

*11 important adverse drug reactions (ADRs) are indicated; note that this list is not comprehensive

ADR, adverse drug reactions; AI, adrenal insufficiency; GI, gastrointestinal; IVIg, intravenous immunoglobulins; MMP, mucous membrane pemphigoid; PDAI, Pemphigus Disease Activity Index; PML, progressive multifocal leukoencephalopathy.

eTABLE 2

Treatment options for pemphigus diseases

Disease/ severity	Treatment	
	Medication*2	Adverse drug reactions*11
Pemphigus vulgaris/ Pemphigus folia- ceus*1 (24)		
Mild (PDAI ≤ 15)	Prednisolone*3 p.o. 1.0–1.5 mg/kg/day or i.v. steroid pulses*4 (level C) ± azathioprine*5 (level C) mycophenoles*6 (level C) dapsone*7 (only for pemphigus foliaceus) (level E)	Hypertension, diabetes, osteoporosis, AI, infections Myelo- and hepatotoxicity, arthralgias, infections GI complaints, infections, (rarely) myelotoxicity Hemolytic anemia, GI complaints
Moderate, severe (PDAI > 15)	Rituximab 2 × 1 g*8 (level B)+ prednisolone*3 p.o. 1.0 mg/kg/day or i.v. steroid pulses*4 (tapering over 3–6 months; level B) ± azathioprine*5 or mycophenols*6 (level C) prednisolone*3 p.o. 1.0–1.5 mg/kg/day or i.v. steroid pulses*4 + azathioprine*5 or mycophenols*6 (level C)	Infusion reaction, infections, reactivation of hepatitis B, PML (ADRs of these drugs are described above)
Refractory to treatment	IVIg*9 (level D) Immunadsorption*10 (level D)	Headache, chills, flushing, fever, hypertension, nausea Hypotension, thrombosis, bradycardia, anaphylaxis, herpes zoster infection
Paraneoplastic pemphigus (2, 3)	Treatment of neoplasm + prednisolone*3 p.o. 0.5–1.0 mg/kg/day or i. v. steroid pulses*4 eventually in combination with rituximab*8, IVIg*9, im- munadsorption*10 (all: level E)	(ADRs of these drugs are described above)
IgA pemphigus (2, 3, e94, e95)	Dapsone*7 and/or acitretine + Prednisolone*3 p.o. 0.5–1.0 mg/kg/day (all: level E)	Hemolytic anemia, GI complaints, teratogenicity Hypertension, diabetes, osteoporosis, AI, infections

*1 German and/or European therapy guidelines are available for these diseases

*2 Level of evidence: level A, meta-analyses of prospective, controlled trials; level B, high-quality prospective, controlled trials; level C, lower-quality prospective controlled trials; level D, major case studies; level E, small case studies, case reports

*3 or prednisolone equivalent

*4 dexamethasone 100 mg/day for three consecutive days, or prednisolone 500–1 000 mg/day, every 3–4 weeks, eventually every 6–8 weeks

*5 2.0–2.5 mg/kg/day with normal thiopurine methyltransferase levels

*6 mycophenolate mofetil (2 g/day), mycophenolate sodium (1 440 mg/day)

*7 1.0–1.5 mg/kg/day with normal glucose 6-phosphate

*8 or biosimilar

*9 2 g/kg for 2–5 days every four weeks; after six months, interval extension (37)

*10 on 3–4 consecutive days with regenerable adsorbers every 3–4 weeks, for 8–16 weeks

*11 not a comprehensive list

ADR, adverse drug reactions; AI, adrenal insufficiency; GI, gastrointestinal; IVIg, intravenous immunoglobulins; PDAI, Pemphigus Disease Activity Index; PML, progressive multifocal leukoencephalopathy.